



INTERNATIONAL JOURNAL OF PHARMACY & LIFE SCIENCES
(Int. J. of Pharm. Life Sci.)

Stroke: Is a major culprit for cerebrovascular disease?

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Abstract

Stroke is an important cause of morbidity and mortality and the second leading cause of dementia worldwide. The main cause of stroke is the intermission of the blood supply to the neurons either due to ischemia or due to bursting of blood vessels. The pathophysiology of the stroke is very complex and is difficult to understand. There are various pathological conditions which provoke stroke included platelet aggregation, mitochondrial dysfunction, hyperhomocysteinemia, glutamate excitotoxicity, various genetic disorders, destruction of endothelial cell wall, atherosclerotic plaque formation, accumulation of bilirubin within the neurons, hyperfibrinogenemia, accumulation of inflammatory mediators like interleukins, chemokines, cytokines, neutrophils, leukocytes etc. results in loss of neuronal functions and neuronal death. The objective of this review is to throw light over the various pathophysiological pathways involves in the evolution of stroke.

Key-Words: Hyperhomocysteinemia, Stroke, Interleukins

Introduction

Stroke is a heterogeneous group of cerebrovascular conditions and is a sudden and devastating illness. However, many people are unaware of its widespread impact [1]. A stroke or “brain attack” occurs when a blood clot blocks the blood flow in a vessel or artery, interrupting blood flow to an area of the brain result damage of brain cells. When brain cells die during a stroke, abilities controlled by that area of the brain are lost. These include functions such as speech, movement, and memory [2, 3]. However, the World Health Organization defines a stroke as “Rapidly developing clinical signs of focal or global disturbance of cerebral function lasting more than 24 hours with no apparent cause other than a vascular origin” [4]. Moreover, Cardiovascular stroke is a broad term used to describe the syndrome in which acute vascular and brain tissue change manifests as one or more neurologic defects that last more than 24 hours, these defect results from either inadequate blood flow or hemorrhage [5]. Researchers observed that stroke is a global health problem. However, it is the second leading cause of death and fourth Leading cause of disability worldwide.

Approximately 20 million people each year will suffer from stroke and of these 5 million will not survive. As in developed countries, stroke is the first leading cause of disability, the second leading cause of dementia [6]. It is also a predisposing factor for epilepsy, falls and depression in developed countries [7]. It can be identified from the symptoms occurs during it include: vertigo, sensory loss, facial numbness, ataxia, dysphagia, dysarthria, ophthalmoplegia, arm and leg paralysis, amnesia, color amnesia, or coma. Stroke patient's suffers from many other complications like sleep problem, confusion, depression, incontinence, atelectasis, pneumonia and swallowing dysfunction that causes aspiration, dehydration or malnutrition. Immobility can lead to thromboembolic disease, reconditioning, sarcopenia, UTIs, pressure ulcer and contractors. Daily functions include the ability to walk, see, feel, remember, think, and speak may be decreased [8-10]. Stroke can be majorly classified into two types including ischemic stroke and hemorrhagic stroke. An ischemic stroke is that kind which includes narrowing or blocking of the blood vessels that leads to the brain's supply due to which the supply of blood to the brain is cut off [11]. The most common cause of ischemic stroke in humans is occlusion of the middle cerebral artery (MCA). It can be thrombotic, embolic, or due to systemic hypoperfusion or to venous thrombosis [12]. Hemorrhagic stroke is the one in which the blood vessels present in the brain or situated near the brain

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bursts that causes bleeding and stopping the oxygen supply to the surrounding areas of the brain [13]. Hypoxia is the major reason for hemorrhagic stroke which caused destruction of the brain tissues and vasculature due to which intracranial pressure is increased resulting in bleeding [14]. This present review article described the pathophysiology of stroke.

Material and Methods

Pathophysiology of Ischemic stroke

Role of inflammatory mediators in acute ischemic stroke: - Astrocyte and glial cells are the building blocks of neurovascular system. These cells act as a bridge between vessels and neurons [15]. As, the major function of these cells is to maintain the homeostasis, therefore, is a crucial mediator of inflammatory responses in case of brain injury. Cerebral ischemia evokes the inflammatory mediators such as Cytokines and chemokine. However, the chemokine and cytokines further involve the enrollment of leukocytes. Moreover, the physiological role of leukocyte involves phagocytosis and regulation of molecular events including the thrombin formation which is further responsible for the cellular changes such as chemotaxis, proliferation and more pronounced cytokine and chemokine release [16, 17]. However, the most pertinent subfamilies of chemokine that would act as a booster in cerebral ischemia are CC and CXC. These two will be the reason for the assessment of neutrophil and monocyte which act as phagocytic cells [18]. Endothelial cells secrete the cell adhesion molecule in response to the inflammatory changes. Now, these cell adhesion molecules further promote the glial cells to secrete chemokine after ischemic stroke [19]. However, chemokine already reside in the brain at very low levels, which includes CX3CL1 (Fractalkine) present in neurons and CXCL12 resides in astrocytes. The major functions of chemokine in the brain are to maintain the homeostasis [20]. After the brain Injury, the level of chemokine goes on increasing which recruits all the inflammatory mediators like TNF – α , IL-1, IL-6 resulting in the activation of ischemic cascade [21]. Moreover, CCL2, CCL5 and CXCL8 are chemokine that will participate in the pathophysiology of stroke [22]. However, CD40 receptors are also responsible for the activation of proinflammatory mediators. The over activation of the CD40 receptors caused the recruitment of cytokines and adhesion molecules (Fig 1) [23].

Role of homocysteine in stroke: - Homocysteine is the amino acid that carries Sulfhydryl group. The demethylation of methionine results in the production of Homocysteine [24]. However, homocysteine is responsible for the various changes include lipid

peroxidation, free radical formation, and Inflammation which further leads to atherosclerosis [25, 26]. The sulfhydryl group is the culprit for all the damages occurred by homocysteine. Sulfhydryl group is responsible for the autoxidation of homocysteine and thiolactone formation that would further lead to atherosclerotic plaque formation [27]. However, hyperhomocysteinemia would lead to the production of peroxynitrite. Moreover, this peroxynitrite stimulates Poly (ADP-ribose) Polymerase (PARP). As PARP stimulates the uncoupling of NO synthase leads the nitration of protein tyrosine residue results in vascular damage and arise oxidative stress. This would result in the imbalance between vasoconstrictor and vasodilator factors [28]. Hyperhomocysteinemia restricts the growth of endothelial cells, decreases the expression of adhesion molecules and leads to the formation of low density lipids (LDL). It also modifies coagulation and fibrinolysis system [29].

Role of ATP and glutamate in stroke: - Brain is the organ which consumes high energy level of the body so it needs high glucose and oxygen. Any restriction in the flow of blood causes brain injury. However, cerebral ischemia leads to ATP depletion in the neurons, glial cells and small vessels which further results in lactic acid accumulation. The accumulated lactic acid provokes the formation of free radicals and increases the release of glutamate [30]. Moreover, reduced levels of ATP inhibit the Na^+/K^+ -ATPase leads to depolarization of neurons and astrocytes. However, on hypoxia astrocyte activates the glycolytic activity permitting the continued uptake of glutamate from the synaptic cleft to the penumbra. [31]. The increased level of glutamate further activates the glutamate N-methyl-d-aspartate (NMDA) receptors. The over activation of glutamate receptors results in excitotoxicity is the condition in which cell dies due to the toxic actions of excitatory amino acids [32, 33]. However, glutamate involved in a number of pathological changes includes impairment of calcium buffering, activation of mitochondrial permeability transition pore and free radical generation [34]. Therefore, excessive release of glutamate causes mitochondrial dysfunction, which further causes the assessment of apoptosis- caspase cascade leads to cell death (Fig. 2) [35, 36].

Role of endothelium in stroke: - The major role of cerebral endothelium is to regulate hemostasis and the formation of blood brain barrier (BBB) [37]. To achieve homeostasis, blood flow brings antithrombotic and anti-inflammatory agent to endothelium, which includes thrombomodulin-protein-C-protein S system, heparin like molecules, tissue factor pathway

inhibitors, tissue plasminogen activator (t-PA), prostacyclin, adenosine diphosphatase, deminase enzyme which removes C-terminal tryptophan from endothelin-1. However, at the time of injury endothelium undergoes considerable changes, i.e. activation of antithrombotic factors and anti-inflammatory mediators. Therefore, leukocyte activates the adenosine receptors [38]. The other changes include activation of platelet activating factors such as factor VIII/von willebrand factor, thromboxane A₂ [39] and the three isoforms of endothelin i.e. ET-1, ET-2, ET-3 which further results in the accumulation of various inflammatory mediators, vasoconstriction and brain hemorrhage [40, 41]. Moreover, decreases the matrix ligands like laminin-1, laminin-5, collagen-1 and fibronectin from the basal lamina results in middle cerebral artery occlusion that further escalate neuronal injury. As, Ischemic insult causes swelling of astrocytes, decreases the consistency of cytoplasmic density and swelling of endothelium therefore, brings middle cerebral artery occlusion. [42]

Role of hepatic encephalopathy in stroke:- Hepatic Encephalopathy causes the accumulation of toxic metabolites in the body like ammonia, urea. Cirrhosis is the condition that causes severe hepatic damage and destruction of hepatocytes [43]. However, intestine absorbed toxic metabolites, but the destructed liver is unable to excrete that from the body which further transported to brain through blood. There it increases the permeability of blood brain barrier. Moreover, auto-regulation of cerebral blood flow is impaired in case of hepatic encephalopathy [44], which results in decreasing the metabolism of oxygen in the brain due to which brain suffers from hypoxia. However, it may also result in decreasing the velocity of blood to middle cerebral artery that provokes cerebral ischemia [45].

Role of cardiac embolism in stroke:- Destruction of cardiac muscles causes the condition called as cardiac dysfunction, which further results in improper functioning of cardiac pumps. Therefore, results in the restriction of the flow of blood due to which thrombin red clot is formed. However, the thrombus fragments are accomplished in the systemic arterial circulation [46]. As, the heart supplies 20% of blood to the brain, therefore, the clot embolized to an intracranial artery. This results in the occlusion of intracranial artery. Moreover, blood applies force to restore the supply of blood to the brain, which further causes dispersion of clot and formed embolus at the different sites of artery [47].

Role of toll like receptors (TLRs) in stroke:- TLRs are the superfamily that mediates innate immune responses. It is Class 1 Membrane Protein which

comprised of ectodomains, transmembrane and intracellular TLR domains. When the ligands bind to TLR that brings conformational changes in the receptor [48]. These changes enrolled the intracellular adaptor proteins such as MyD88, TRIF, TIRAP or TRAM all of it causes the activation of NF- κ B (Nuclear Factor Kappa B). The ligands which activate the receptor include heme, fibrinogen, high mobility group protein B1, hyaluronan, oxidized low density lipoprotein, ayloid β [49]. TLRs mediate its effect by the activation of two signaling pathways, which is activated by the adaptor protein i.e. MyD88 and TRIF. The activation of MyD88 regulates the action of pro-inflammatory cytokine genes and activation of NF- κ B. However, the activation of TRIF includes TLR3 and TLR4 mediated the pathway, which further include the enrollment of IRF-3 results in stimulation of IFN- β [50]. Therefore, IFN- β involves the recruitment of macrophage infiltration. However, the activation of NF- κ B recruits TLR2 and TLR4 which evoke brain ischemia. TLR4 pathway further leads to the activation of AP-1 (activating protein 1) which is dimeric protein. However, this dimeric protein consists of members of Jun, FOS and α -Feroprotein. Furthermore, MMPs includes MMP2, MMP3, MMP9, MMP12 and Protease activation enrolled t-PA [51]. However, AP-1 also includes the assessment of mitogen activated protein kinase like c-Jun-N-terminal kinase (JNK), p38, extracellular signal regulated kinases (ERK) [52].

Role of monogenic diseases in stroke:- It had been observed that about 5% of stroke cases are reported due to the mutation in the genes. Gene-gene interaction or gene-environment interaction being the major reason for this mutation. These genetic factors modify the target organs and therefore, become the risk factors that are responsible for the stroke [53]. Following are the some mutational changes that are responsible for the stroke

Familial Hemiplegic Migraine:- The mutation of CACNA1A gene is responsible for childhood stroke [54]. However, CACNA1A gene encoding Alpha 1A sub-unit of Voltage Gated Calcium Channels [55].

Cerebral Autosomal Dominant (CADASIL) Arteriopathy with subcortical infarct and Leukoencephalopathy:- Mutation of NOTCH3 gene is the major reason for the evolution of this disease which further evokes stroke with the passage of time [56]. However, this is the disease of cerebral small vessels, which further causes the vascular cognitive impairment. Moreover, Basal Ganglia and Temporal Lobe suffered from ischemia due to this disease and developed ischemic lesions in these areas [57].

Fabry disease: - It is X-linked congenital lysosomal storage disorder [58]. However, impairment in α -Galactosidase that causes the accumulation of glycosphingolipids in the vascular endothelial cells, kidneys, heart, neurons which further results in tissue damage [59].

Role of renin angiotensin aldosterone system in stroke: - Angiotensin converting enzyme (ACE) is a membrane bound enzyme which is very important for the conversion of angiotensinogen to angiotensin I (ANG I) and angiotensin II (ANG II). However, ANG I and ANG II play the crucial role in the development of hypertension, atherosclerosis, and cardiovascular diseases [60]. ACE gene is located on chromosome 17q23 and also contains 26 exons. However, 287 bp located in intron 16 at ACE gene is responsible for the polymorphism by addition and deletion. Furthermore, this polymorphism leads to ischemia, which further developed infarction [61].

Role of sickle cell anemia and stroke:- Sickle cell anemia is the genetic disorder which plays the heterogeneous role in the development of stroke. Sickle red blood cells stick to the vascular endothelium, which creates hydrostatic pressure and shear stress [62]. However, these blood cells first of all bind to thrombospondin, VCAM (vascular adhesion molecule), P-Selectin which further creates the interaction between sickle red blood cells and vascular endothelium results in vascular occlusion [63]. Moreover, sickle cell anemia leads to an infarction in childhood and hemorrhage in adults. Furthermore, these cells also activate the accumulation of NF- κ B, endothelin, and leukocytes that further worsen the occlusion results in a stroke [64].

Role of ischemic effect in stroke: - Decrease in the blood flow toward the brain due to many reasons like rupturing of arterioles and parenchymal hemorrhage, which abruptly decreases the blood pressure or intracerebral hemorrhage that bring a reduction in the perihematomal blood flow [65]. However, the perihematomal blood reduction becomes the major cause, for the accumulation of proteins within the neurons. The proteins such as: heat shock protein 105, L-lactate dehydrogenase A, Peroxiredoxin 1 (PRDX1), glutathione-S-transferase P, α -enolase, pyruvate dehydrogenase E1, fructose-bisphosphate aldolase C, 6-phosphofructo kinase type C, catalase, lactoglylutathione lyase, cytoplasmic enzymes of glycolysis [66]. Moreover, the ischemic condition of neurons leads to the reduction of enolase level caused hypoxia [67].

Role of septin-5 protein, synaptojanin-1 protein and dynamic related protein-1 (DRP-1) in stroke:-

Septin 5 proteins are localized in the pre-synaptic terminal that holds negative effect on the release of neurotransmitters. It had been observed that social isolation brings the stressful conditions due to which septin- protein level decreased within the stratum which further results in ischemia of the human brain [68]. Moreover, DRP-1 is important for the outgrowth of axons by binding with the calcium channels as well as it also helps in the formation of mitochondrial outer membrane. Mutation or upregulation of DRP-1 leads to the fragmentation of mitochondria, further results in evolution of apoptosis and cell death [69, 70]. However, synaptojanin-1 protein also called phosphoinositide phosphate is distributed in the nerve terminals. Its major function is the maintenance of the amount of phosphoinositol-4, 5-biphosphate within the synaptic membrane. However, the increase in its level during ischemia causes destruction of synaptic membrane [71].

Role of hyperbilirubinemia in stroke:- The byproduct of heme metabolism is bilirubin. However, when elevated level of bilirubin is concentrated within the neurons, it produced the toxic substances like urea, ammonia in the neurons which further results in the production of oxidative stress. Moreover, increased concentration of bilirubin also caused the aggregation of halogenated hydrocarbon which acts as mediator of oxidative stress [72].

Role of microRNA (miRNA) in stroke:- It is endogenous small non-coding regulatory RNA molecules. It damages the messenger RNA (mRNA) through binding with their coding targets. It plays very important role in the pathogenesis of stroke. However, miRNA shows functions like neural cell survival, impaired regulation of neurovascular integrity, stroke mediated inflammation [73]. The miRNA that participated in the evolution of stroke includes miR-124, miR-9, miR-219, miR132. These miRNA modifies the blood brain barrier by accumulating cytokines and thus causes the breakdown of blood brain barrier [74]. Moreover, miRNA involved in endothelial functions, vascular functions, erythropoiesis, angiogenesis, neural functions, hypoxia. However, circulating miRNA is the end product of microvesicle secretion which is involved in RNA-binding Agronature (Ago) proteins [75]. Necrotic cells secreted Ago proteins into the systemic circulation from the cytoplasm of the cell through destruction of cellular membranes. Therefore, it caused middle cerebral artery occlusion. In case of autoimmune encephalomyelitis, the plasma concentration of miRNA-124 is increased and miRNA-9 is decreased, thus facilitates the neuroinflammation during the stroke. [76]

Role of lipoprotein associated phospholipase A2 (Lp-PLA2) in stroke:- Lp-PLA2 is a calcium independent phospholipase and is also called as Platelet-activating factor acetyl hydrolase (PAF-AH). However, it is involved in the cleavage of oxidized Phosphatidylcholine and thus formed lysophosphatidyl choline (lyso-PC) and oxidized non-esterified acids [77]. Therefore, it participates in the activation of proinflammatory mediators. Moreover, Lp-PLA2 is encoded by PLA2G7 gene, which is localized on chromosome 6p21-p12. However, the polymorphism of V2797 which is present in exon 9 of PLA2G7 gene associated with the development of coronary diseases, stroke, and carotid atherosclerosis [78].

Role of stromelysin-1 in stroke:- It is a matrix metalloproteinase that further regulate extracellular matrix in case of tissue injury. Therefore, it gives rise to the growth of atherosclerotic plaque. In its gene sequence the promoter of the gene contains 600bp which have 6 adenosine (6A) whereas the other genes contain only 5 Adenosine (5A). However, 5A allele shows more pronounced activity than that of 6A allele. Therefore, 6A allele contains lower levels of stromelysin in arterial walls results in the accumulation of matrix within the cells, which further increases the thickness of carotid artery walls [79].

Role of hyperfibrinogenemia in stroke: - It is the condition of elevated plasma fibrinogen levels. Hyperfibrinogenemia leads to the impairment of the blood coagulation system which increases the blood viscosity and decreases the flow of blood. Therefore, it promotes the platelet aggregation and adhesion to the endothelial cells [80]. However, fibrinogen is a dimeric glycoprotein contains 3 pairs of polypeptide chains, i.e. two strands of A α , B β and γ . Moreover, these six polypeptide chains construct Hexamer (A α B β γ)². Furthermore, these polypeptide chains, i.e. A α , B β and γ are encoded by the different genes alpha fibrinogen gene (FGA), beta fibrinogen gene (FGB), gamma fibrinogen gene (FGG) respectively. However, single nucleotide polymorphism (SNPs) of B β effects plasma fibrinogen levels, therefore, B β limits the production and maturation of fibrinogen. Moreover, SNPs in FGB at -148C/T located in FGB promoter gene observed that person with T allele have higher fibrinogen level than C allele [81].

Conclusion

As stroke is a global health problem, therefore, it is being very important to know about the pathological pathways, further it helps in the treatment of stroke. Ischemia causes the accumulation of inflammatory mediator's results in phagocytosis, thrombin formation and oedema. Moreover, the deficiency of folic acid and

vitamin B12 leads to hyperhomocysteinemia which further results in lipid peroxidation, free radical formation, and endothelial dysfunction. However, endothelial dysfunction causes imbalance between vasodilators and vasoconstrictors agents. Excessive secretion of glutamate due to depletion of ATP causes over activation of NMDA glutamate receptor further results in mitochondrial destruction. Therefore, mitochondria start releasing caspase-3, -7 and 9 which results in apoptotic cell death. Sometimes, the genetic disorders also provoke a stroke like familial hemiplegic migraine, cerebral autosomal dominant arteriopathy with subcortical infarct and leukoencephalopathy, fabry disease, sickle cell anemia. Moreover, the mutation of ACE gene also brings the condition which favors the evolution of stroke. However, activation of TLR with the specific ligands causes the activation of TLR-2 and TLR-4. Moreover, accumulation of bilirubin within the neurons results in the production of toxic substances and oxidative stress. MicroRNA leads to the necrotic cell death, miR-124 and miR-9 are the major microRNA which is involved in the pathogenesis of stroke. This article has compiled the pathophysiological roles of various types of diseases, mediators, gene and proteins in stroke.

Acknowledgement

We wish to express our gratefulness to Dr. S.L. Harikumar (Honourable Director-RBIP), Sr. Gurdinder Singh Bahra Ji (Honourable Chancellor), Dr. S. K. Bansal (Honourable Vice Chancellor) of Rayat and Bahra University Mohali (Punjab) for their praiseworthy inspiration, platform and constant support for the completion of this study.

References

1. Mergenthaler P, and Meisel A. (2012). Do Stroke models model Stroke?. *Disease Models and Mechanisms*, 5: 718-725
2. Kumar A.S, and Kumar Reddy TS. (2012). A review on Brain Attack. *International Journal of Pharmacology and Toxicology*, 2(1): 44-54.
3. Sacco RL, Kasner SE, Broderick JP, Caplan LR, Culebras A, Elkind MSV, George MG, Hamdan AD, Higashida RT, Hoh BL, Janis LS, Kase CS, Kleindorfer DO, Lee JM, Moaseley ME, Peterson ED, Turan TN, Valderrama AL, and Vinters HV. (2013). An updated definition of Stroke for the 21st Century: A Statement for Healthcare professionals from the American Heart Society/ American Stroke Association. *Stroke*, 44: 1524-4628.
4. Eleftheriou D. (2012). Endothelial Injury and Repairs in childhood Arterial Ischemic Stroke.

- Department of Paediatric Rheumatology, Institute of Childhealth, University College London, 3-292.
5. Jeffrey S. (2009). Low income countries bear biggest burden of stroke. *Medscape Medical News*.
 6. Taylor FC, and Kumar S. (2012). Stroke in India Factsheet. *South Asian Network for Chronic Disease*, IIPH Hyderabad, Pulic Health Foundation of India: 1-25
 7. Fisher M, and Bo N. (2011). 1st Global Conference on Healthy Life Styles and Non-Communicable disease control. *Moscow*.
 8. Humphries SE, and Morgan C. (2004). Genetic risk factors for stroke and carotid atherosclerosis insights into pathophysiology from candidate gene approaches. *The Lancet Neurology*, 3: 227-336.
 9. Davenport RJ, Dennis MS, Wellwood I, and Warlow CP. (1996). Complications After Acute Stroke. *Stroke*, 27(3): 415-420
 10. O' Collins V.E, Donnam G.A, Macleod M.R, and Howells D.W (2013). Animal Model of Stroke versus Clinical Stroke : Comparison of Infarct Size, Cause, Location, Study Design and Efficacy of Experimental Therapies. *Animal Model for Study of Human Diseases*.
 11. Dutta T, and Bajaj MM. (2014). Neurophysical and Neurochemical study of Acute Brain attack with special reference to matrix string theory: Anti Bis Mechanism and their role in prevention of neuronal entropy enhancement. *Asian Journal Of Medical Sciences And Clinical Research*, 1(1): 1-12.
 12. Feigin V, Lawes C, Bennet D, Barkr Cello S, and Parag v. (2009). Worldwide stroke incidence and early case fatality in 56 population based studies: A systemic review. *Lancet Neurology*, 8(4): 355-369.
 13. Gungd BM, Jagtap PN, Ingale VB, and Patil RY. (2013). Stroke : A Brain Attack. *IOSR Journal of Pharmacy*, 3(8): 01-23.
 14. Bamio DE. (2015). Hearing Disorders in the Stroke. *Handbook of Clinical Neurology*, 129: 1-25.
 15. Jessen K.R. (2004). Cells in Focus Glial Cells. *The International Journal of Biochemistry and Cell Biology*, 36: 1861-1867.
 16. Downey G.P. (1997). Effect of Mechanical deformation on structure and function of Polymorphonuclear Leukocytes. *Journal of Applied Physiology*, 82: 1395-1396.
 17. Bouchard B.A, and Tracy P.B. (2003). The participation of Leukocytes in coagulant Reactions. *Journal of Thrombosis and Haemostasis*, 1: 464-489.
 18. Wook Oh J, Schwiebert LM, and Benveniste EN. (1999). Cytokine regulation of CC and CXC Chemokine expression by human astrocyte. *Journal of Neurovirology*, 5: 82-94.
 19. Albelda SM, Smith CW, and Ward PA. (1994). Adhesion molecules and inflammatory injury. *FASEB Journal*, 8(8): 504-512.
 20. Garcia-Berrocso T, Giralto D, Victorillobart, Bustamante A, Penalba A, Flores A, Ribo M, Molina C.A, Rosell A, and Montaner J (2014). Chemokines after Human Ischemic Stroke: From Neurovascular Unit to Blood using Protein Arrays. *Translation Alproteomics*, 3: 1-9
 21. Lehmann J, Hartig W, Seidel A, Fuldner C, Hobohm C, Grosche J, Krueger M, and Michalski D. (2014). Inflammatory Cell Recruitment After Experimental Thromboembolic Stroke in Rats. *Neuroscience*, 0306-4522.
 22. Luo Y, Zhou YQ, Xiao W, Liang Z, Dai J, Weng X, and Ulu X. (2014). IL-3 ameliorates Ischemic Brain Injury in Experimental Stroke through promoting Th2 response and suppressing Th17 responses. *Brain Research*, 12: 005.
 23. Ying M, Wang HX, Liu Y, Peng GG, Wang XM, Zhang B, Hua Wu B, and Yu JM. (2013). Single Nucleotide Polymorphism of CD40 in the 5'-untranslated region is associated with Ischemic Stroke. *Gene*.
 24. Paul G, and Alam F. (2015). Role of Homocysteine in the development of Cardiovascular disease. *Nutrition Journal*, 14:6
 25. Bayir A. (2015). The Effects of Vitamin B12 and Folic Acid Deficiencies on Stroke, and Vitamin B12 and Folic Acid Supplement. *Foods and Dietary Supplements in the Prevention and Treatment of Disease*.
 26. Manolescu B.N, Oprea E, Farcasanu IC, Berteanu M, and Cercasov C. (2010). Homocysteine and vitamin therapy in stroke prevention and treatment: a review. *ACTA ABP Biochimica Polonica*, 57: 467-477
 27. Karolczak K, and Olas B. (2009). Mechanism of Action of Homocysteine and Its Thiolactone in Hemostasis System. *Physiological Research*, 58: 623-633.

28. Nanetti L, Vignini A, Raffaelli F, Giulietti A, Bartolini M, Perozzi C, Silvestrini C, Provinciali L, and Mazzanti L. (2013). Homocysteine and oxidative stress in acute stroke. *Advances in Bioscience and Biotechnology*, 4: 15-23
29. Singh RB, Mengi SM, Xu YJ, Arnega AS, and Dhalla NS. (2002) Pathogenesis of atherosclerosis: A multifactorial process. *Experimental and Clinical Cardiology*, 7(1): 40-53
30. Wang Z, Xiao J, Xie S, Zhao D, Liu X, Zhang J, Yuan Y, and Huang Y. (2012). Evaluation of Cerebral Oxygen Metabolism and Blood Flow in Stroke like episodes of MELAS. *Journal of Neurological Sciences*, 323: 173-177.
31. Brambilla R, Couch Y, and Lambertsen KL. (2013). The effect of Stroke on immune functions. *Molecular and Cellular Neuroscience*, 53: 26-33.
32. Graham SH, and Hickey RW. (2002). Molecular Pathophysiology Of Stroke. *Neuropsychopharmacology: The Fifth Generation of Progress*, 1317-1326.
33. Lucke-Wold BP, Logsdon AF, Turner RC, Rosen CL, and Huber JD. (2014). Aging, the metabolic syndrome and ischemic stroke: Redefining the approach for studying the Blood Brain Barrier in a complex neurological disease. *Advances in Pharmacology*, 71: 1054-3589.
34. D.J. Michelson, and S. Ashval. (2003). The Pathophysiology of Stroke in Mitochondrial Disorder. *Mitochondrion*, 4: 665-674.
35. Breton RR and Rodriguez JCG. Excitotoxicity and Oxidative Stress in Acute Ischemic Stroke”, *www.intechopen.com*.
36. Yin KJ, Hamblin M, and Chen YE. (2014). Non-Coding RNAs in cerebral Endothelial Pathophysiology: Emerging roles in Stroke. *Neurochemistry International*, 0197-0186.
37. Sandoval K.E., and Witt K.A. (2014). Blood Brain Barrier tight function permeability and Ischemic Stroke. *Neurobiology Disorder*, 32: 200-219.
38. Woywodt A, Gerdes S, Ahl B, Erdbruegger U, Haubitz M, and Weissenborn K. (2012). Circulating Endothelial cells and Stroke: Influence of Stroke Subtype and Changes during the Course of Disease. *Journal of Stroke and Cerebrovascular Disease*, 21 (6): 452-458.
39. Eggers AE. (2006). Factor XII (Hageman Factor) is a missing link between stress and hypercoagulability and plays an important role in the pathophysiology of ischemic stroke. *Medical Hypothesis*, 67: 1065-1071.
40. Paczkowska E, Janouska MG, Bajer-Czajkowska A, Machalinska A, Utianouski P, Rybicka M, Klos P, Dziedziejko V, Safranow K, and Nowacki P, B. (2013). Increased Circulating Endothelial Progenitor Cells in patients with Hemorrhagic and Ischemic Stroke: The role of Endothelin-1. *Journal of Neurological Sciences*, 325: 90-99.
41. Kaundal RK, Deshpande TA, Gulati A, and Sharma SS. (2012). Targeting endothelin receptors for pharmacotherapy of ischemic stroke: Current scenario and future perspectives. *Drug Discovery Today*, 17: 1359-6446.
42. Zoppo GJD, and Hallenbeck JM. (2000). Advances in the Vascular Pathophysiology of Ischemic Stroke. *Thrombosis Research*, 98: V73-V81.
43. Husseini NE, Kaskar O, and Goldstein LB. (2014). Chronic Kidney Disease and Stroke. *Advances in Chronic Kidney Disease*, 21 (6): 500-508.
44. Rodriguez G, Testa R, Celle G, Gris A, Marengo S, Nobili F, Novellone G, and Rosadini G. (1987). Reduction of Cerebral Blood flow in subclinical hepatic Encephalopathy and its correlation with plasma free Tryptophan. *Journal of Cerebral Blood Flow and Metabolism*, 7: 768-772.
45. Yamamoto Y, Nishiyama Y, Katsura K, Yamazaki M, and Katayama Y. (2011). Hepatic Encephalopathy with Reversible Focal Neurologic Signs Resembling Acute Stroke: Case Report. *Journal of Stroke and Cerebrovascular Diseases*, 20 (4): 377-380.
46. Rote WE, Nedelman MA, Mu DX, Manley PJ, Weisman H, Cunningham MR, and Lucchesi BR. (1994). Chimeric TE3 prevents carotid artery thrombosis in Cynomolgus Monkeys. *Stroke*, 25: 1223-1233.
47. Radlidis LS, Zindaki MG, Vikelis M, Kaliva K, Papadopoulos C, and Kremastinos DT, (2009). Elevated soluble intercellular adhesion molecule-1 levels are associated with poor short term prognosis in middle-aged patients with acute ischemic stroke. *International Journal of Cardiology*, 132: 216-220.

48. Macrez R, Ali C, Toutiris O, Mouff B, Defer G, Dimagl U, and Vivien D. (2011). Stroke and Immune System: From Pathophysiology to new therapeutic strategies. *Lancet Neural*, 10: 471-480.
49. Fang H, Wang PF, Zhou Y, Wang YC, and Yang QW. (2013). Toll-Like receptor 4 Signalling in intracerebral Hemorrhage-induced inflammation and injury. *Journal of Neuroinflammation*, 10: 27
50. Ownes CED, and Crack PJ. (2010). Neural Injury Following Stroke: are Toll-Like Receptors the link between the Immune System and the CNS?. *British Journal of Pharmacology*, 160: 1872-1888.
51. Tang SC, Arumugam TY, Langru XU, Cheng A, Mughal MR, Gyu Jo D, Lathia JD, Siller DA, Chigurupati S, Ouyang X, Magnus T, Comandola S, and Mattson MP. (2007). Pivotal Role for Neuronal Toll-Like Receptor in Ischemic Brain Injury and functional deficits. *Proceedings of the National Academy of Sciences of the United States of America*, 104: 13798-13803.
52. Asai C, and Asai H. (2014). Involvement of Toll-Like Receptor in Ischemic Stroke Induced Neuronal damage. *Journal of Neurological disorders and Stroke*, 2 (2): 1051.
53. Lalouschek W, Schillinger M, Hsieh K, Endler G, Tentschert S, Lang W, Cheng S and Mannhalter C. (2005). Matched Case Control Study on factor V Leiden and the prothrombin G20210A Mutation in patients with Ischemic Stroke/ Transient Ischemic Attack up to the age of 60 years. *American Heart Association*, 36: 1405-1409.
54. Landi A, Marotta N, Mancarella C, Marruzzo D, Salvati M, and Delfini R. (2011). Basal ganglia stroke due to mild head trauma in pediatric age - clinical and therapeutic management: a case report and 10 year literature review. *Italian Journal of Pediatrics*, 37: 2.
55. Carenno O, Corominas R, Serra SA, Sintas C, Castillu NF, Vila-Pueyo M, Toma C, Gene GG, Pons R, Llana M, Sobrido MJ, Grinberg D, Valverde MA, Fernandez-Fernandez JM, Macaya A, and Cormand B. (2013). Screening of CACNA1A and ATP1A2 genes in hemiplegic migraine: clinical, genetic, and functional studies. *Mol Genet Genomic Med*, 1 (4): 206-222.
56. Stojanov D, Grozdanovic D, Petrovic S, Benedeto-Stojanov D, Stefanovic I, Stojanovic N, and Llic DN. (2014). De novo mutation in the NOTCH3 gene causing CADASIL. *Bosn J Basic Med Sci*, 14(1): 48-50.
57. Bohlega S. (2011). Novel mutation of the notch3 gene in Arabic family with CADASIL. *Neurology International*, 3: e6.
58. Yu P, Cui Y, Cai W, Wu H, Xiao X, Shao Q, Ma L, Guo S, Wu N, Jin ZB, Wang Y, Cai T, Sun ZS, and Qu J. (2015). Lysosomal storage disease in the brain: mutations of the β -mannosidase gene identified in autosomal dominant nystagmus. *Genetics in Medicine*, 10: 1038.
59. Clarke T.R. Joe. (2007). Narrative Review: Fabry Disease. *Annals of Internal Medicine*, 146: 425-433.
60. Munshi A, Das S, and Kaul S. (2014). Genetic determinants in Ischemic Stroke Subtype; Seven year findings and a Review. *Gene*
61. Terni E, Giannini N, Brondi M, Montano N, Baonucelli U, and Mancuso M. (2015). Genetic of Ischemic Stroke in young adults. *BBA Clinical*, 3: 96-106.
62. Driss A, Asare KO, Hibbert JM, Gee Be, Adamkiewicz Tv, Stiles JK. (2009). Sickle cell Disease in the post Genomic era: A Monogenic Disease with a polygenic phenotype. *Genomics Insights*, 2: 23-48.
63. Switzer JA, Hers DC, Nichols FT, and Adams RJ. (2006). Pathophysiology and Treatment of Stroke in Sickle Cell Diseases: Present and future. *Lancet Neurology*, 5: 501-512.
64. Marano M, Quattrocchi C, Annibali O, Avvisati G, and Di Lazzaro V. (2014). Recruitment Large Volume Silent Stroke in Sickle Cell Diseases. *Journal of Stroke and Cerebrovascular Diseases*, 23 (10): 453-455.
65. Tayal A.H. (2007). Quantitative Perihematomal Blood Flow in Spontaneous Intracerebral Hemorrhage Predicts In-Hospital Functional Outcome. *Stroke*, 38:319-324.
66. Jaremko K.M., Roetling J.C., Chen L., and Regan R.F. (2011). Accelerated Hemolysis and Neurotoxicity in Neuron-Glia-Blood Clot Co-cultures. *J Neurochem*, 114(4): 1063-1073.
67. Mulder M, and Geocadin R.G. (2013) Hypoxic-Ischemic Encephalopathy in Adults. *Neurocritical Care Society Review Course*.

68. Zholumbetov E. (2011). The role of Septin 5 in Exocytosis. *Department of Biochemistry, University of Toronto*
69. Tian Y, Li B, Shi WZ, Chang MZ, Zhang GJ, Di ZL, and Liu Y. (2014). Dynamin-Related Protein 1 Inhibitors Protect against Ischemic Toxicity through Attenuating Mitochondrial Ca²⁺ Uptake from Endoplasmic Reticulum Store in PC12 Cells. *International Journal of Molecular Sciences*, 15: 3172-3185.
70. Zhang L, Zhijie H, Zhang Q, Wu Y, Yang X, Niu W, Hu Y, and Jia J. (2014). Exercise Pretreatment Promotes Mitochondrial Dynamic Protein OPA1 Expression after Cerebral Ischemia in Rats. *International Journal of Molecular Sciences*, 15: 4453-4463.
71. Ren C, Guingab-Cagmat J, Koleissy F, Zoltewicz S, Mondello S, Gao M, Hafeez A, Li N, Ging X, Larner SF, Anagli J, Hayes RL, Ji X, and Ding Y. (2014). A Neuroproteomic and systems biology analysis of rat brain post intracerebral hemorrhagic stroke. *Brain Research Bulletin*, 102: 46-56.
72. Pineda S, Bang Y, Saver JL, Starkman S, Yun SW, Liebeskind DS, Kim D, Ali LK, Shah SH, and Ovbiagele B. (2008). Association of Serum Bilirubin with Ischemic Stroke Outcomes. *Journal of Stroke and Cerebrovascular Disease*, 17 (3): 147-153.
73. Chen J, Venkat P, Zacharek A, and Chopp M. (2014). Neurorestorative Therapy for Stroke. *Frontiers in Human Neuroscience*, 8: 382.
74. Wang Y, Wang Y, and Yang GY. (2013). MicroRNAs in Cerebral Ischemia. *Stroke Research and Treatment*, 276540.
75. Liu Y, Zhang J, Han R, Liu H, Sun D, and Liu. (2015). Downregulation of serum brain specific MicroRNA is associated with inflammation and infarct volume in acute Ischemic Stroke. *Journal of Clinical Neurosciencce*, 22: 291-295.
76. Wang C, Bingyaun, Cheng B, Chen J, and Bai B. (2014). Neuroprotection of microRNA in neurological disorders (Review). *Biomedical Reports*, 2(5): 611-619.
77. Iribarren C, Gross MD, Darbinian JA, Jacobs DR, Sidney S, and Loria CM. (2005). Association of Lipoprotein-Associated Phospholipase A2 Mass and Activity With Calcified Coronary Plaque in Young Adults The CARDIA Study. *Arterioscler Thromb Vasc Biol.*, 25: 216-221.
78. Liu X, Zhu RX, Tian YL, Li Q, Li L, Deng SM, and He ZY. (2014). Association of PLA2G7 gene polymorphisms with Ischemic Stroke in Northern Chinese Han population. *Clinical Biochemistry*, 47: 404-408.
79. Humphries SE, and Morgan L. (2004). Genetic risk factors for stroke: Insights into pathophysiology from candidate gene approaches. *International Congress Series*, 1262: 482-485.
80. Swarowska M, Polczak A, Pera J, Mrowiec AK, Slowik A, and Dziedzic T. (2014). Hyperfibrinogenemia predicts long-term risk of death after ischemic stroke. *Journal of Thrombosis and Thrombolysis*, 38: 517-521.
81. Inran I, Ponpon RL, Idjradinata, Achmad TH, Maskoen A, Wibowo S, and Harapan H. (2015). Association of fibrinogen promoter gene polymorphism (-148C/T), Hyperfibrinogenemia and Ischemic Stroke in young adult patients. *The Egyptian Journal of Medical Human Genetics*, 16: 11-17.

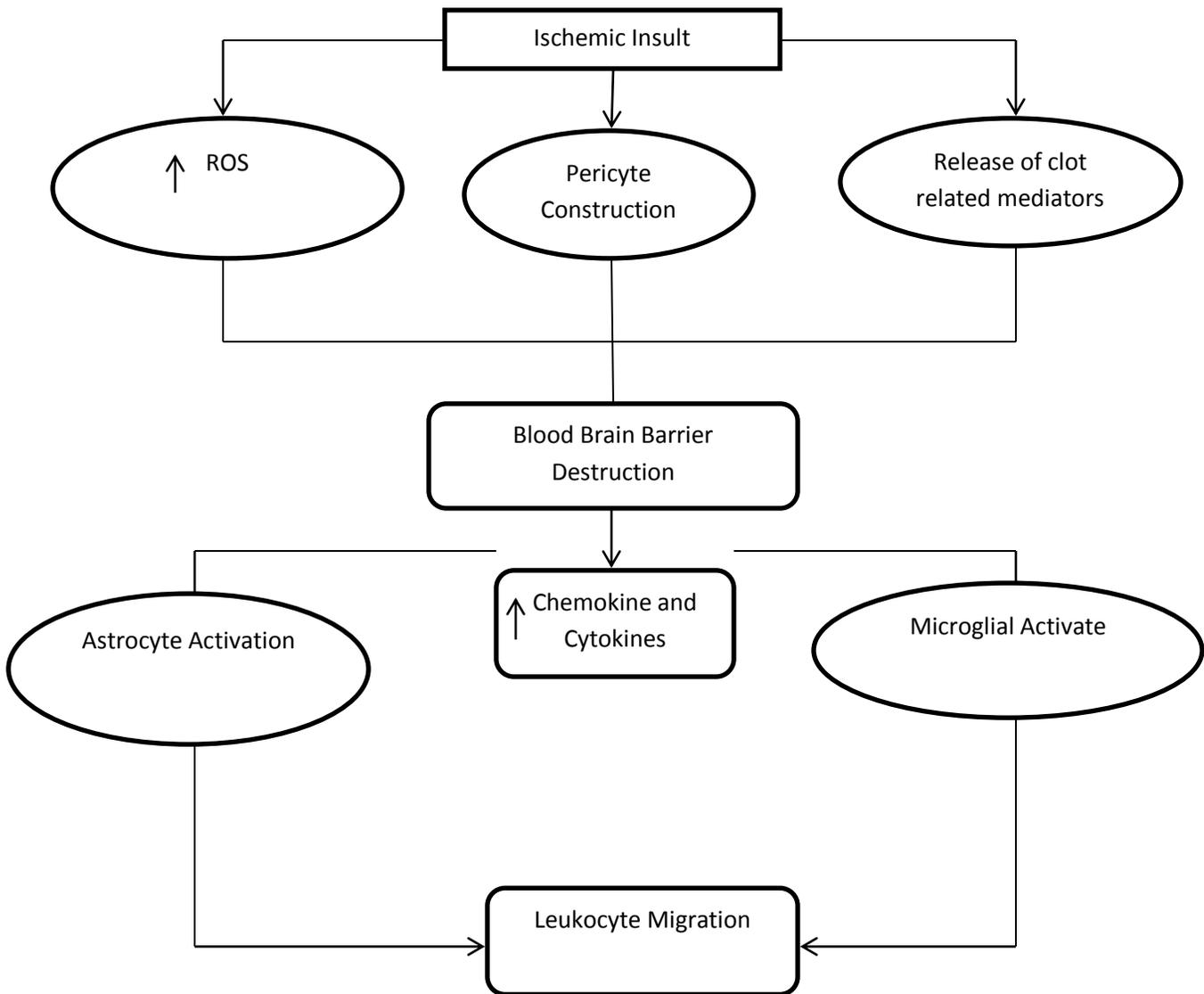


Fig. 1: The role of inflammatory mediators resulting leukocyte migration.

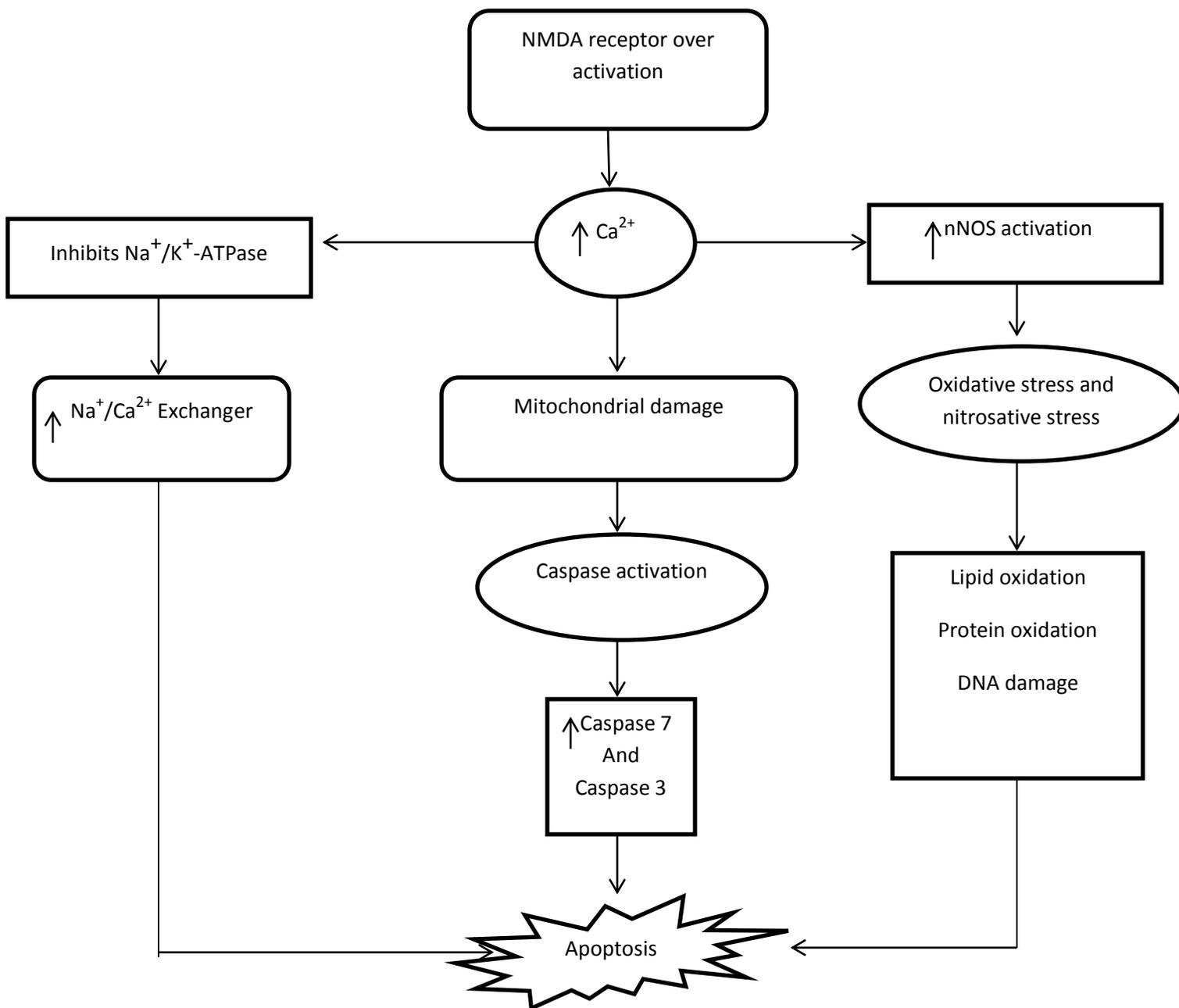


Fig. 2: Figure indicates the over-activation of NMDA glutamate receptors lead to apoptotic cell death

How to cite this article

Singh G., Kaur R. and Karikumar S.L. (2015). Stroke: Is a major culprit for cerebrovascular disease?. *Int. J. Pharm. Life Sci.*, 6(7):4595-4605.

Source of Support: Nil; Conflict of Interest: None declared

Received: 10.06.15; Revised: 26.06.15; Accepted: 20.07.15